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Summary

Improving the rate sperm retrieval rate of patients afflicted with nonobstructive azoospermia is a critical issue. In this study, I designed an *in vivo* RNA interference experiment by using a vector plasmid that expressed siRNAs against *Numb* (numb homolog (Drosophila)) and *Numbl* (numb homolog (Drosophila)-like), which are reported to be necessary for spermiogenesis in mouse; the vector was transfected into rat testis *in vivo* and the influence of vector transfection was evaluated. I analyzed the expression levels and localization of *Numb* and *Numbl* in normal rat testes. Furthermore, I performed a loss-of-function analysis using a gene-knockdown (KD) approach *in vivo*.

10 Normal 8-week-old rats were divided into 4 groups based on the gene KD: Group A: *Numb* KD; Group B: *Numbl* KD; Group C: *Numb/Numbl* co-KD; and Group D: negative control. Histological evaluations were performed at 7 and 14 days after gene transfer. The expression patterns of *Numb* and *Numbl* were distinct. No differences in the proportion of germ cells were detected between the 4 groups at 7 days after

15 transfection. However, at 14 days after knocking down *Numb* and *Numbl* expression, the number of spermatids decreased substantially in the Group-A rats, and the number of elongated spermatids decreased markedly in the rats in Groups B and C. No remarkable damage was observed in vector-transfected testes. Thus, siRNA-expressing vectors were

safely transfected into testes and they expressed siRNAs *in vivo* and caused minimal damage. Furthermore, I found that *Numb* and *Numbl* regulate the differentiation of spermatids during rat spermatogenesis. It is expected that these observations to form the basis for the future development of gene therapy or molecule-targeted therapies for male

5 infertility in which the Notch signaling system is modified using *Numb* and *Numbl*.